



American Society of Hematology 2021 L Street NW, Suite 900, Washington, DC 20036 Phone: 202-776-0544 | Fax 202-776-0545 editorial@hematology.org

What is the future of immunotherapy in MM?

Tracking no: BLD-2019-004176-CR1

Leo Rasche (University Hospital Wuerzburg, Germany) Michael Hudecek (Universitaetsklinikum Wuerzburg, Germany) Hermann Einsele (Universitätsklinikum Würzburg, Germany)

Abstract:

The treatment of Multiple Myeloma (MM) is currently being redefined by humoral and cellular immunotherapies. For decades, there was limited believe in immune-based anti-MM therapy due to the moderate graft-versus-myeloma effect of allogeneic stem cell transplantation. Today, monoclonal antibodies are the new backbone of anti-MM therapy, and T-cell therapies targeting BCMA are emerging as the most potent single agents for MM treatment. Herein, we present our assessment and vision for MM immunotherapy in the short- and mid-term future.

Conflict of interest: COI declared - see note

COI notes: L.R. participated in scientific advisory boards for Janssen, Celgene/BMS, GSK, Sanofi, and has received research support from Skyline Dx . M.H. participated in scientific advisory boards for Janssen and Celgene/BMS; and is listed as an inventor on patent applications related to CAR technologies that have been filed by the University of Würzburg. H.E. Scientific Advisory Board: Janssen, Celgene/BMS, Amgen, Novartis, Takeda. Research Support: Janssen, Celgene/BMS, Amgen, Novartis. Honorarium: Janssen, Celgene/BMS, Amgen, Novartis, Takeda

Preprint server: No;

Author contributions and disclosures: L.R., M.H. and H.E. wrote and approved the manuscript.

Non-author contributions and disclosures: No;

Agreement to Share Publication-Related Data and Data Sharing Statement:

Clinical trial registration information (if any):

What is the future of immunotherapy in MM?

Rasche L^{1,2}, Hudecek M¹, Einsele H¹

- ¹ Medizinische Klinik und Poliklinik II, Universitätsklinikum Würzburg, Würzburg, Germany
- ² Mildred Scheel Early Career Center, Universitätsklinikum Würzburg, Würzburg, Germany

Corresponding author:

Prof. Dr. Hermann Einsele Universitätsklinikum Würzburg Medizinische Klinik und Poliklinik II Oberdürrbacherstrasse 6 97080 Würzburg Germany Fon: +49 (0) 931 201 40001 Fax: +49 (0) 931 201 640001 Email: <u>Einsele H@ukw.de</u>

ABSTRACT

The treatment of Multiple Myeloma (MM) is currently being redefined by humoral and cellular immunotherapies. For decades, there was limited believe in immune-based anti-MM therapy due to the moderate graft-versus-myeloma effect of allogeneic stem cell transplantation. Today, monoclonal antibodies are the new backbone of anti-MM therapy, and T-cell therapies targeting BCMA are emerging as the most potent single agents for MM treatment. Herein, we present our assessment and vision for MM immunotherapy in the short- and mid-term future.

The future is now: antibody immunotherapy is the new backbone of MM therapy

After the approval of combination therapies with monoclonal antibodies (mAb) for patients with relapsed Multiple Myeloma (MM), the anti-CD38 mAb daratumumab (Dara) has become the new backbone of first-line therapy in transplant-eligible and -ineligible MM patients. In the CASSIOPEIA trial, a quadruple-regimen of Dara, bortezomib, thalidomide and dexamethasone (Dara-VTD) accomplished an overall response rate (ORR) and a progression free survival (PFS) of 93% and 93% vs. 90% and 85% in the control arm at 18-months¹, respectively, setting a new benchmark for efficacy in induction therapy. Minimal residual disease (MRD)-negativity was achieved in 64% of patients versus 44% in the VTD control arm, suggesting improved overall survival (OS) with longer follow-up. Based on these results, the U.S. (FDA) and EU (EMA) regulatory agencies approved Dara-VTD in early 2020.

Likewise, the induction regimen Dara-VRD, comprising lenalidomide instead of thalidomide, achieved compelling results in the GRIFFIN phase II study with PFS and OS rates of even \geq 95% at 24 months². Notably, PFS rates were not significantly different between Dara-VRD and VRD, but there was a marked difference in MRD negativity in the Dara arm (51.0% vs 20.4% at 22 months), which will likely translate into better PFS with longer follow-up ³. The phase 3 study PERSEUS of Dara-VRD versus VRD has completed enrollment and results are expected to be reported in 2022. In transplant-ineligible MM patients, combination therapies that comprise Dara as backbone, i.e. Dara-VMP⁴ and Dara-Rd⁵, have recently been approved. Dara s.c. injection⁶ has been approved lately and accordingly, Dara-VMP and Dara-Rd can be considered the new standard for transplant-ineligible patients with newly diagnosed (ND)MM.

Building on the successful clinical use of Dara, alternative anti-CD38 antibodies are entering the stage with isatuximab (Isa) being the most clinically advanced candidate. In the IKARIA trial, the triple-regimen of isatuximab, pomalidomide randomized and dexamethasone was superior to the control arm of pomalidomide and dexamethasone in relapsed refractory (RR)MM patients, with a PFS of 11.5 months and 6.5 months, respectively⁷. Based on this study, the FDA approved this regimen in March 2020 and we anticipate approval by EMA to occur in 2020 as well. In analogy to the PERSEUS trial, the GMMG HD7 trial evaluates induction therapy with Isa-VRD versus VRD in a randomized fashion. It is likely that both anti-CD38 antibodies - Dara and Isa - will be approved in combination regimen for use in first-line therapy. Numerous additional regimen with anti-CD38 antibodies, e.g. in combination with carfilzomib^{8,9} or traditional chemotherapy¹⁰ are being evaluated and will enrich the therapeutic armamentarium to achieve disease control in RRMM patients.

The primary mode of action of Dara and Isa is distinct however, the activity of both mAbs is in part dependent on the density of CD38 molecules on the myeloma cell membrane¹¹. Accordingly, strategies to increase CD38-expression on MM cells, e.g. by epigenetic modulation are being investigated^{12,13}. Of note, Dara also depletes CD38+ immunosuppressive cells, which is associated with an increase of cytotoxic T cells, potentially contributing to the activity of this antibody¹⁴. An unresolved question is whether prior therapy with an anti-CD38 mAb impacts on the subsequent efficacy of T-cell redirecting therapies. Because CD38 is expressed on activated T cells (and to a lesser extent on resting T cells and on NK cells), anti-CD38 mAbs may lead to pertubations in T cell composition and may interfere with the mode-of-action of chimeric antigen receptor (CAR) T-cell therapy and

T-cell engaging bispecific antibodies (bsAb). Careful investigations are warranted to determine the optimal sequence and timing of anti-CD38 therapy and T-cell redirecting therapies.

The role of elotuzumab (Elo), an anti-SLAMF7 mAb, in MM therapy is less clear. The anti-MM potency of Elo as a single agent is rather limited and therefore, Elo has been evaluated in combination with immune-modulating agents to augment activity. Recently, final data from the ELOQUENT-1 trial, that evaluated Elo, lenalidomide and dexamethasone (Elo-RD) versus RD in transplant-ineligible NDMM patients, did not report additive activity of Elo¹⁵. However, in the relapse setting, Elo-RD was superior to the RD control arm¹⁶. Furthermore, Elo in combination with pomalidomide and dexamethasone exerted significant clinical activity in the ELOQUENT-3 study with 40% of RRMM patients being in remission at 2 years¹⁷. Appreciating that Elo is well tolerated and has a favorable safety profile, this antibody may also play a role in the setting of maintenance therapy. Currently, Elo is evaluated in a quadruple induction and consolidation regimen in transplant-eligible patients with NDMM (Elo-VRd - HD6 trial, Elo-KRd - DSMMXVII trial). Notably, T-cells expressing a SLAMF7-CAR with a targeting domain derived from Elo are substantially more potent against MM than Elo in pre-clinical models in vitro and in vivo¹⁸ and therefore, the results of phase I/IIA clinical trials with SLAMF7-CAR T-cells (CARAMBA and MELANI-01) are eagerly awaited.

A new future begins: the dawn of T cell redirecting therapies

Chimeric antigen receptor T-cells

T-cell redirecting therapies with gene-engineered CAR T-cells and T-cell engaging bsAb are currently the most exciting new developments in cancer immunotherapy and will change the treatment paradigm for MM (**Figure 1**). Idecabtagene-vicleucel (Ide-cel; bb2121) is a BCMA-CAR T-cell therapy that reported an ORR of 85%, with 45% of patients achieving complete response (CR) in heavily pretreated RRMM in a dose-escalating phase 1 study¹⁹. The pivotal phase 2 KarMMa study with Ide-cel has fully recruited and initial results were presented at the 2020 ASCO meeting showing an ORR of 73% and a median PFS of 8.6 months, and both increased with higher dose²⁰. Thus, we are expecting FDA approval of Ide-cel for RRMM in 2020, which will constitute a landmark as the first gene-engineered T-cell therapy in MM.

LCAR-B38M (also known as JNJ-68284528) is another compelling BCMA-CAR T-cell product currently under investigation in phase I/II trials²¹. Interim results from the CARTITUDE-1 study showed an ORR of 100% with 76% stringent CRs for heavily pretreated RRMM patients²². Several other BCMA-CAR T-cell products are being investigated in phase 1 studies and collectively, the data support the notion that BCMA-CAR T-cells are highly effective and probably are the most potent single agent available in the RR disease setting^{19,23,24}. However, duration of response is limited in these trials and the majority of patients ultimately relapse. In contrast to CD19 CAR T- cells in Non-Hodgkin lymphoma ²⁵, there is no survival plateau in MM so far.

Overall, BCMA-CAR T-cell therapy displayed a favorable safety profile with lower incidence of cytokine release syndrome and neurotoxicity compared to CD19-CAR T-cell therapy in B-cell leukemia and -lymphoma^{19,26}. However, the clinical experience with BCMA-CAR T-cells has also exposed several challenges associated with targeting this antigen, and potential mechanisms of relapse or resistance include antigen down-regulation or even loss ²⁷ in a

small subset of patients, as well as limited persistence of BCMA-CAR T-cells, and limited fitness of T-cells in heavily pre-treated patients^{19,21,23,24,28}. Several strategies are pursued to address these challenges, including the use of gamma-secretase inhibitors to enhance BCMA molecule density on MM cells and to reduce the amount of soluble BCMA in serum²⁹; refined CAR T-cell manufacturing protocols to increase fitness, e.g. in the presence of PI3K inhibitors³⁰; the use of CAR products with defined T-cell subset composition and humanized targeting domains to reduce immunogenicity and to promote engraftment and in vivo expansion^{29,31,32}.

Antibody drug conjugates

A subset of MM patients with significant comorbidities may be less able to tolerate the potential toxicity associated with T-cell redirecting therapies and for this patient population, BCMA-specific antibody drug conjugates (ADC) may be an alternative. A lead candidate in this class of compounds is belantamab mafadotin. Belantamab mafadotin eliminates MM cells by releasing the cytotoxic agent auristatin F and through antibody-dependent cellular cytotoxicity (ADCC). Belantamab mafadotin is administered i.v. every 3 weeks and is well tolerated except of corneal toxicity, which occurs in >70% of patients³³. The ORR with belantamab mafadotin was around 30% in a phase 2 trial addressing patients refractory to Dara, IMiDs and proteasome inhibitors³⁴. PFS was 2.8 months and 3.9 months in the 2.5mg/kg and 3.4 mg/kg dose group, respectively. In responding patients, PFS was not reached and 8.4 months in the 2.5 mg/kg and 3.4 mg/kg dose group, respectively. Intriguingly, belantamab mafadotin's mode-of-action is independent of T-cell fitness and accordingly, this ADC is given consideration as one of the few remaining therapeutic options for patients that experience relapse after CAR T-cell therapy³⁶.

The future ahead: a race between BCMA-targeting agents to first-line therapy?

T-cell engaging bispecific antibodies

T-cell engaging bsAbs are another way to harness the power of a cellular immune response to combat MM (**Figure 1**). A first proof-of-concept for bsAbs in MM was recently provided with AMG420, a CD3xBCMA bispecific T-cell engager (BiTE) construct. At the dose of 400 µg/d, the response rate was 70% including 50% MRD-negative complete responses, with some responses lasting >1 year³⁷. Recently, a pilot study with the asymmetric CD3xBCMA bsAb CC-93269³⁸ reported an 89% ORR and a 44% CR rate at the highest dose of 10 mg in heavily pre-treated MM patients³⁹. Teclistamab and REGN5458 are two other BCMA/CD3 bispecific antibodies with promising preliminary results in early clinical trials ^{40,41}.

A practical advantage of bsAbs over CAR T-cells is that they are 'off-the-shelf' products and can be administered repeatedly to sustain the therapeutic pressure against MM. A current focus in the field is to determine the most active bsAb compound that we anticipate will advance to pivotal trials within the next 1-3 years, and to derive detailed insights into potential mechanisms of resistance to bsAb therapy due to interference from soluble BCMA protein, BCMA down-regulation, as well as humoral and cellular immune responses against synthetic bsAb constructs⁴².

We anticipate that T-cell redirecting therapies targeting BCMA (CART, bsAb) will rapidly move forward from late-stage RRMM to earlier treatment lines, and even to first-line therapy in difficult-to-treat MM patients. A key assumption supporting this strategy is that the fitness of endogenous T cells at early disease stages is higher compared to later disease stages, when patents have received multiple rounds of cytotoxic therapy⁴³. While data to support this hypothesis are still emerging in MM⁴⁴, we have recently shown in lymphoma that patient T cells acquire functional defects after chemotherapy, which impacts on activity of bsAbs⁴⁵.

A challenge for establishing BCMA-CAR T-cells and bsAbs in first-line therapy is the requirement to demonstrate superiority over standard-of-care, where Dara-based combinations have now set a high bar in efficacy and safety. Accordingly, several studies focus on subgroups of MM patients with suboptimal response to and outcome after conventional and Dara-based combination regimen, because these patients may particularly benefit from CAR T-cell and bsAb therapy. The randomized KarMMa 3 study evaluates Idecel versus standard-of-care in RRMM patients previously treated with Dara, an IMiD and a proteasome inhibitor, who have received at least 2 but not more than 4 prior regimen. The CARTITUDE-4 study investigates LCAR-B38M in patients with 1-3 prior lines of therapy, and pre-exposed to proteasome inhibitors and resistant to lenalidomide. Of note, there is limited safety data on BCMA CAR T-cell therapy in the transplant ineligible MM patient population. Similarly, there is a strategy to implement the ADC belantamab mafadotin earlier in MM treatment, e.g. the DREAMM 6 study evaluates belantamab mafadotin in combination with RD or VD in second line. The DREAMM 9 study will evaluate belantamab mafadotin together with standard of care as induction therapy for transplant eligible NDMM patients.

Key requirement: biomarkers to guide patient selection and choice of immunotherapy

Suboptimal response to and early relapse after induction therapy and high-dose chemotherapy with subsequent autologous stem cell transplantation⁴⁶ is another approach to define MM patients that may particularly benefit from CAR T-cell therapy as an element of first-line treatment. An alternative strategy is to identify high-risk MM patients at primary diagnosis using molecular markers, which is not without challenges due to the inter- and intra-patient tumor heterogeneity in $MM^{47,48}$. We consider several markers as being potentially suitable for identifying high-risk patients for inclusion into CAR T-cell and bsAb therapy studies, including R-ISS stage 3^{49} , TP53 double-hit event^{50,51}, gene-expression profiling-defined high-risk status^{52,53}, and the presence of extramedullary MM disease^{54,55}. The most recent FISH-based MM risk classifier is another promising tool for patient stratification⁵⁶. We would consider an OS of \geq 3 years in this MM patient subgroup a breakthrough that now seems accomplishable in the immunotherapy era.

MRD is another key marker defining a subgroup of MM patients with suboptimal response and outcome. The number of MRD-positive patients is significant and constitutes around 40% of NDMM patients after high-dose chemotherapy and autologous stem cell transplantation¹. For these patients, the goal of administering CAR T-cell therapy is conversion to MRD-negativity with ensuing long-term disease-free survival and even cure. It is important to note that eradication of MM cells in the MRD setting is a prerequisite for cure. Recent data suggest that a primary mode-of-action of CAR T-cells against hematologic cancer cells is the induction of apoptosis⁵⁷, and it remains to be determined whether metabolically-inactive MM cells in MRD-positive patients can be sufficiently eliminated. Double-hit TP53 lesions are found in 4% of NDMM patients⁵⁸, increase in frequency in the course of disease^{59,60} and provide another mechanism of apoptosis resistance. A recent study exposed that impaired FAS receptor signaling is associated with failure of CD19-CAR T-cell therapy in acute leukemia⁶¹. Noteworthy, genes involved in apoptosis induction are frequently altered due to mutations and chromosomal aberrations in MM^{62,63}.

Burning questions and outlook: The future is bright for immunotherapy in MM!

There are several burning questions centered around *i*. identifying biomarkers that predict outcome and identify patients that have the highest chance to benefit from T-cell redirecting therapies; *ii*. identifying the optimal single antigen or combination of antigens for T-cell redirecting therapies to consistently induce durable complete remissions; and *iii*. determine whether eventually, MM can be cured in a relevant subset (or even the majority) of patients (**Table 1**). At our institution, we have treated MM patients with biallelic TP53 inactivation and very aggressive disease and observed rapid and deep responses after BCMA-CAR T-cell therapy. This is in line with data from e.g. the KarMMa study, where the majority of patients with high-risk cytogenetics and extramedullary disease responded to CAR T-cell treatment¹⁹. These data suggest that current algorithms for staging and risk assessment in MM ought to be adapted in the immunotherapy era.

There is a rich pipeline of novel CAR T-cell products, bsAbs, and trispecific antibodies⁶⁴ that target alternative antigens including e.g. SLAMF7¹⁸, CD44v6⁶⁵ and GPRC5D⁶⁶, as well as multi-specific CAR T-cells⁶⁷. Indeed, it will be important to determine whether targeting two (or more) antigens can exert more therapeutic pressure to counteract down-regulation of a single antigen and clonal evolution of MM cells. In another approach, NK cells are used as effector cells and CAR-modified NK cells as well as bispecific killer cell engager (BiKE) targeting MM antigens showed encouraging results in preclinical studies ^{68,69}. A recent report on a Phase 1/2 trial evaluating CD19-CAR NK cells in patients with non-Hodgkin's lymphoma and chronic lymphocytic leukemia showed a high rate of initial responses and a favorable toxicity profile ⁷⁰. We have recently demonstrated that advanced microscopy techniques can be used to identify and monitor target antigens on MM cells to inform therapeutic decisions⁷¹. Significant efforts are undertaken to simplify the manufacturing and logistics around CAR Tcell therapy involving virus-free gene-transfer, automated point-of-care production and allogeneic cell products^{72,73}. With these developments we are confident that the role of immunotherapy in MM will be manifested and the prospect of a chemo-free free, yet curative treatment for the majority of MM patients can become 'real' within the next decade.

Acknowledgements

The authors are supported by German Research Foundation (Deutsche Forschungsgemeinschaft; project number 324392634, TRR 221 to M.H. and H.E.); the German Ministry for Education and Research (Bundesministerium für Bildung und Forschung; project IMMUNOQUANT to M.H. and H.E.); the European Union's Horizon 2020 research and innovation programme under grant agreements No. 733297 (EURE-CART to M.H. and H.E.) & No. 754658 (CARAMBA to M.H. and H.E.); the Myeloma Crowd Research Initiative (to M.H. and H.E.); the patient advocacy group 'Hilfe im Kampf gegen den Krebs e.V.', Würzburg, Germany and 'Forschung hilft' - Stiftung zur Förderung der Krebsforschung an der Universität Würzburg (L.R., M.H.and H.E.). L.R. is supported by German Cancer Aid (Deutsche Krebshilfe e.V.) as a fellow in the Mildred Scheel Early Career Center Würzburg (Mildred Scheel Nachwuchzentrum).

Author contributions

Conflicts-of-interest

L.R. participated in scientific advisory boards for Janssen, Celgene/BMS, GSK, Sanofi, and has received research support from Skyline Dx .

M.H. participated in scientific advisory boards for Janssen and Celgene/BMS; and is listed as an inventor on patent applications related to CAR technologies that have been filed by the University of Würzburg.

H.E. Scientific Advisory Board: Janssen, Celgene/BMS, Amgen, Novartis, Takeda. Research Support: Janssen, Celgene/BMS, Amgen, Novartis. Honorarium: Janssen, Celgene/BMS, Amgen, Novartis, Takeda

References

1. Moreau P, Attal M, Hulin C, et al. Bortezomib, thalidomide, and dexamethasone with or without daratumumab before and after autologous stem-cell transplantation for newly diagnosed multiple myeloma (CASSIOPEIA): a randomised, open-label, phase 3 study. *Lancet.* 2019;394(10192):29-38.

2. Voorhees P, Kaufman JL, Laubach J, et al. Daratumumab plus Lenalidomide, Bortezomib & Dexamethasone Improves Depth of Response in Transplant-eligible Newly Diagnosed Multiple Myeloma: GRIFFIN. *Clinical Lymphoma Myeloma & Leukemia*. 2019;19(10):E353-E354.

3. Voorhees PM, Kaufman JL, Laubach JP, et al. Daratumumab, Lenalidomide, Bortezomib, & Dexamethasone for Transplant-eligible Newly Diagnosed Multiple Myeloma: GRIFFIN. *Blood.* 2020.

4. Mateos MV, Dimopoulos MA, Cavo M, et al. Daratumumab plus Bortezomib, Melphalan, and Prednisone for Untreated Myeloma. *N Engl J Med.* 2018;378(6):518-528.

5. Facon T, Kumar S, Plesner T, et al. Daratumumab plus Lenalidomide and Dexamethasone for Untreated Myeloma. *N Engl J Med.* 2019;380(22):2104-2115.

6. Usmani SZ, Nahi H, Mateos MV, et al. Subcutaneous delivery of daratumumab in relapsed or refractory multiple myeloma. *Blood*. 2019;134(8):668-677.

7. Attal M, Richardson PG, Rajkumar SV, et al. Isatuximab plus pomalidomide and low-dose dexamethasone versus pomalidomide and low-dose dexamethasone in patients with relapsed and refractory multiple myeloma (ICARIA-MM): a randomised, multicentre, open-label, phase 3 study. *Lancet.* 2019;394(10214):2096-2107.

8. Zhou X, Fluchter P, Nickel K, et al. Carfilzomib Based Treatment Strategies in the Management of Relapsed/Refractory Multiple Myeloma with Extramedullary Disease. *Cancers (Basel)*. 2020;12(4).

9. Chari A, Martinez-Lopez J, Mateos MV, et al. Daratumumab plus carfilzomib and dexamethasone in patients with relapsed or refractory multiple myeloma. *Blood*. 2019;134(5):421-431.

10. Zhou X, Steinhardt MJ, Grathwohl D, et al. Multiagent therapy with pomalidomide, bortezomib, doxorubicin, dexamethasone, and daratumumab ("Pom-PAD-Dara") in relapsed/refractory multiple myeloma. *Cancer Med.* 2020.

11. Moreno L, Perez C, Zabaleta A, et al. The Mechanism of Action of the Anti-CD38 Monoclonal Antibody Isatuximab in Multiple Myeloma. *Clin Cancer Res.* 2019;25(10):3176-3187.

12. Garcia-Guerrero E, Gogishvili T, Danhof S, et al. Panobinostat induces CD38 upregulation and augments the antimyeloma efficacy of daratumumab. *Blood*. 2017;129(25):3386-3388.

13. Garcia-Guerrero E, Gotz R, Doose S, et al. Upregulation of CD38 expression on multiple myeloma cells by novel HDAC6 inhibitors is a class effect and augments the efficacy of daratumumab. *Leukemia*. 2020.

14. Krejcik J, Casneuf T, Nijhof IS, et al. Daratumumab depletes CD38+ immune regulatory cells, promotes T-cell expansion, and skews T-cell repertoire in multiple myeloma. *Blood.* 2016;128(3):384-394.

15. BMS. <u>https://news.bms.com/press-release/corporatefinancial-news/bristol-myers-</u> squibb-reports-primary-results-eloquent-1-study-; 2020.

16. Lonial S, Dimopoulos M, Palumbo A, et al. Elotuzumab Therapy for Relapsed or Refractory Multiple Myeloma. *N Engl J Med.* 2015;373(7):621-631.

17. Dimopoulos MA, Dytfeld D, Grosicki S, et al. Elotuzumab, Pomalidomide, and Dexamethasone for Relapsed/Refractory Multiple Myeloma: Efficacy After Additional Follow-Up of the ELOQUENT-3 Study. *Clinical Lymphoma Myeloma & Leukemia*. 2019;19(10):E164-E165.

18. Gogishvili T, Danhof S, Prommersberger S, et al. SLAMF7-CAR T cells eliminate myeloma and confer selective fratricide of SLAMF7(+) normal lymphocytes. *Blood*. 2017;130(26):2838-2847.

19. Raje N, Berdeja J, Lin Y, et al. Anti-BCMA CAR T-Cell Therapy bb2121 in Relapsed or Refractory Multiple Myeloma. *N Engl J Med.* 2019;380(18):1726-1737.

20. Munshi NC. Idecabtagene vicleucel (ide-cel; bb2121), a BCMA-targeted CAR T-cell therapy, in patients with relapsed and refractory multiple myeloma (RRMM): Initial KarMMa results. 2020 ASCO: J Clin Oncol 38: 2020 (suppl; abstr 8503); 2020.

21. Zhao WH, Liu J, Wang BY, et al. A phase 1, open-label study of LCAR-B38M, a chimeric antigen receptor T cell therapy directed against B cell maturation antigen, in patients with relapsed or refractory multiple myeloma. *J Hematol Oncol.* 2018;11(1):141.

22. Berdeja JG. Update of CARTITUDE-1: A phase lb/II study of JNJ-4528, a B-cell maturation antigen (BCMA)-directed CAR-T-cell therapy, in relapsed/refractory multiple myeloma. ASCO: J Clin Oncol 38: 2020 (suppl; abstr 8505); 2020.

23. Cohen AD, Garfall AL, Stadtmauer EA, et al. B cell maturation antigen-specific CAR T cells are clinically active in multiple myeloma. *J Clin Invest*. 2019;129(6):2210-2221.

24. Brudno JN, Maric I, Hartman SD, et al. T Cells Genetically Modified to Express an Anti-B-Cell Maturation Antigen Chimeric Antigen Receptor Cause Remissions of Poor-Prognosis Relapsed Multiple Myeloma. *J Clin Oncol.* 2018;36(22):2267-2280.

25. Wang M, Munoz J, Goy A, et al. KTE-X19 CAR T-Cell Therapy in Relapsed or Refractory Mantle-Cell Lymphoma. *N Engl J Med*. 2020;382(14):1331-1342.

26. Moreau P, Sonneveld P, Boccadoro M, et al. Chimeric antigen receptor T-cell therapy for multiple myeloma: a consensus statement from The European Myeloma Network. *Haematologica*. 2019;104(12):2358-2360.

27. Da-Via M. Biallelic deletion of chromosome 16p encompassing the BCMA locus as a tumor intrinsic resistance mechanism to BCMA directed CAR T in Multiple Myeloma. EHA. Frankfurt: EHA Library. Rasche L. 06/12/20; 294800; EP883; 2020.

28. Madduri D, Usmani SZ, Jagannath S, et al. Results from CARTITUDE-1: A Phase 1b/2 Study of JNJ-4528, a CAR-T Cell Therapy Directed Against B-Cell Maturation Antigen (BCMA), in Patients with Relapsed and/or Refractory Multiple Myeloma (R/R MM). *Blood*. 2019;134.

29. Pont MJ, Hill T, Cole GO, et al. gamma-Secretase inhibition increases efficacy of BCMA-specific chimeric antigen receptor T cells in multiple myeloma. *Blood.* 2019;134(19):1585-1597.

30. D'Agostino M, Raje N. Anti-BCMA CAR T-cell therapy in multiple myeloma: can we do better? *Leukemia*. 2020;34(1):21-34.

31. Maude SL, Barrett DM, Ambrose DE, et al. Efficacy and Safety of Humanized Chimeric Antigen Receptor (CAR)-Modified T Cells Targeting CD19 in Children with Relapsed/Refractory ALL. *Blood*. 2015;126(23).

32. Sommermeyer D, Hudecek M, Kosasih PL, et al. Chimeric antigen receptor-modified T cells derived from defined CD8+ and CD4+ subsets confer superior antitumor reactivity in vivo. *Leukemia*. 2016;30(2):492-500.

33. Popat R, Warcel D, O'Nions J, et al. Characterisation of response and corneal events with extended follow-up after belantamab mafodotin (GSK2857916) monotherapy for patients

with relapsed multiple myeloma: a case series from the first-time-in-human clinical trial. *Haematologica*. 2020.

34. Lonial S, Lee HC, Badros A, et al. Belantamab mafodotin for relapsed or refractory multiple myeloma (DREAMM-2): a two-arm, randomised, open-label, phase 2 study. *Lancet Oncology*. 2020;21(2):207-221.

35. Lonial S. Pivotal DREAMM-2 study: Single-agent belantamab mafodotin (GSK2857916) in patients with relapsed/refractory multiple myeloma (RRMM) refractory to proteasome inhibitors (PIs), immunomodulatory agents, and refractory and/or intolerant to anti-CD38 monoclonal antibodies (mAbs). ASCO: J Clin Oncol 38: 2020 (suppl; abstr 8536); 2020.

36. Cohen AD, Garfall AL, Dogan A, et al. Serial treatment of relapsed/refractory multiple myeloma with different BCMA-targeting therapies. *Blood Adv.* 2019;3(16):2487-2490.

37. Topp MS, Duell J, Zugmaier G, et al. Evaluation of AMG 420, an anti-BCMA bispecific T-cell engager (BiTE) immunotherapy, in R/R multiple myeloma (MM) patients: Updated results of a first-in-human (FIH) phase I dose escalation study. *Journal of Clinical Oncology*. 2019;37(15).

38. Seckinger A, Delgado JA, Moser S, et al. Target Expression, Generation, Preclinical Activity, and Pharmacokinetics of the BCMA-T Cell Bispecific Antibody EM801 for Multiple Myeloma Treatment. *Cancer Cell*. 2017;31(3):396-410.

39. Costa LJ, Wong SW, Bermudez A, et al. First Clinical Study of the B-Cell Maturation Antigen (BCMA) 2+1 T Cell Engager (TCE) CC-93269 in Patients (Pts) with

Relapsed/Refractory Multiple Myeloma (RRMM): Interim Results of a Phase 1 Multicenter Trial. *Blood.* 2019;134.

40. Usmani SZ. Phase I study of teclistamab, a humanized B-cell maturation antigen (BCMA) x CD3 bispecific antibody, in relapsed/refractory multiple myeloma (R/R MM). ASCO: DOI: 10.1200/JCO.2020.38.15_suppl.100 Journal of Clinical Oncology 38, no. 15_suppl (May 20, 2020) 100-100.; 2020.

41. Dillillo DJ. REGN5458, a Bispecific BCMAxCD3 T Cell Engaging Antibody, Demonstrates Robust In Vitro and In Vivo Anti-Tumor Efficacy in Multiple Myeloma Models, Comparable to That of BCMA CAR T Cells. ASH 2019: Blood (2018) 132 (Supplement 1): 1944; 2019.

42. Rader C. Bispecific antibodies in cancer immunotherapy. *Curr Opin Biotechnol.* 2019;65:9-16.

43. Garfall AL, Dancy EK, Cohen AD, et al. T-cell phenotypes associated with effective CAR T-cell therapy in postinduction vs relapsed multiple myeloma. *Blood Adv*. 2019;3(19):2812-2815.

44. Danhof S, Schreder M, Knop S, et al. Expression of programmed death-1 on lymphocytes in myeloma patients is lowered during lenalidomide maintenance. *Haematologica*. 2018;103(3):e126-e129.

45. Duell J, Lukic DS, Karg M, et al. Functionally Defective T Cells After Chemotherapy of B-Cell Malignancies Can Be Activated by the Tetravalent Bispecific CD19/CD3 Antibody AFM11. *J Immunother*. 2019;42(5):180-188.

46. Kumar SK, Dispenzieri A, Fraser R, et al. Early relapse after autologous hematopoietic cell transplantation remains a poor prognostic factor in multiple myeloma but outcomes have improved over time. *Leukemia*. 2018;32(4):986-995.

47. Rasche L, Chavan SS, Stephens OW, et al. Spatial genomic heterogeneity in multiple myeloma revealed by multi-region sequencing. *Nat Commun.* 2017;8(1):268.

48. Rasche L, Kortum KM, Raab MS, Weinhold N. The Impact of Tumor Heterogeneity on Diagnostics and Novel Therapeutic Strategies in Multiple Myeloma. *Int J Mol Sci.* 2019;20(5).

49. Palumbo A, Avet-Loiseau H, Oliva S, et al. Revised International Staging System for Multiple Myeloma: A Report From International Myeloma Working Group. *J Clin Oncol.* 2015;33(26):2863-2869.

50. Walker BA, Mavrommatis K, Wardell CP, et al. A high-risk, Double-Hit, group of newly diagnosed myeloma identified by genomic analysis. *Leukemia*. 2018.

51. Shah V, Johnson DC, Sherborne AL, et al. Subclonal TP53 copy number is associated with prognosis in multiple myeloma. *Blood.* 2018;132(23):2465-2469.

52. Shaughnessy JD, Zhan F, Burington BE, et al. A validated gene expression model of high-risk multiple myeloma is defined by deregulated expression of genes mapping to chromosome 1. *Blood*. 2007;109(6):2276-2284.

53. Shah V, Sherborne AL, Johnson DC, et al. Predicting ultrahigh risk multiple myeloma by molecular profiling: an analysis of newly diagnosed transplant eligible myeloma XI trial patients. *Leukemia*. 2020.

54. Usmani SZ, Heuck C, Mitchell A, et al. Extramedullary disease portends poor prognosis in multiple myeloma and is over-represented in high-risk disease even in the era of novel agents. *Haematologica*. 2012;97(11):1761-1767.

55. Rasche L, Angtuaco EJ, Alpe TL, et al. The presence of large focal lesions is a strong independent prognostic factor in multiple myeloma. *Blood*. 2018;132(1):59-66.

56. Perrot A, Lauwers-Cances V, Tournay E, et al. Development and Validation of a Cytogenetic Prognostic Index Predicting Survival in Multiple Myeloma. *J Clin Oncol.* 2019;37(19):1657-1665.

57. Messmer MN, Snyder AG, Oberst A. Comparing the effects of different cell death programs in tumor progression and immunotherapy. *Cell Death Differ*. 2019;26(1):115-129.
58. Walker BA, Mavrommatis K, Wardell CP, et al. A high-risk, Double-Hit, group of newly diagnosed myeloma identified by genomic analysis. *Leukemia*. 2019;33(1):159-170.

59. Weinhold N, Ashby C, Rasche L, et al. Clonal selection and double-hit events involving tumor suppressor genes underlie relapse in myeloma. *Blood*. 2016;128(13):1735-1744.

60. Chavan SS, He J, Tytarenko R, et al. Bi-allelic inactivation is more prevalent at relapse in multiple myeloma, identifying RB1 as an independent prognostic marker. *Blood Cancer J*. 2017;7(2):e535.

61. Singh N, Lee YG, Shestova O, et al. Impaired Death Receptor Signaling in Leukemia Causes Antigen-Independent Resistance by Inducing CAR T-cell Dysfunction. *Cancer Discov*. 2020;10(4):552-567.

62. Stein CK, Pawlyn C, Chavan S, et al. The varied distribution and impact of RAS codon and other key DNA alterations across the translocation cyclin D subgroups in multiple myeloma. *Oncotarget*. 2017;8(17):27854-27867.

63. Gomez-Bougie P, Amiot M. Apoptotic machinery diversity in multiple myeloma molecular subtypes. *Front Immunol.* 2013;4:467.

64. Garfall AL, June CH. Trispecific antibodies offer a third way forward for anticancer immunotherapy. *Nature*. 2019;575(7783):450-451.

65. Casucci M, di Robilant BN, Falcone L, et al. CD44v6-targeted T cells mediate potent antitumor effects against acute myeloid leukemia and multiple myeloma. *Blood.* 2013;122(20):3461-3472.

66. Pillarisetti K, Edavettal S, Mendonca M, et al. A T-cell-redirecting bispecific G-proteincoupled receptor class 5 member D x CD3 antibody to treat multiple myeloma. *Blood*. 2020;135(15):1232-1243.

 Zah E, Nam E, Bhuvan V, et al. Systematically optimized BCMA/CS1 bispecific CAR-T cells robustly control heterogeneous multiple myeloma. *Nat Commun.* 2020;11(1):2283.
 Chan WK, Kang S, Youssef Y, et al. A CS1-NKG2D Bispecific Antibody Collectively Activates Cytolytic Immune Cells against Multiple Myeloma. *Cancer Immunol Res.*

2018;6(7):776-787.

69. Chu J, Deng Y, Benson DM, et al. CS1-specific chimeric antigen receptor (CAR)engineered natural killer cells enhance in vitro and in vivo antitumor activity against human multiple myeloma. *Leukemia*. 2014;28(4):917-927.

70. Liu E, Marin D, Banerjee P, et al. Use of CAR-Transduced Natural Killer Cells in CD19-Positive Lymphoid Tumors. *N Engl J Med*. 2020;382(6):545-553.

71. Nerreter T, Letschert S, Gotz R, et al. Super-resolution microscopy reveals ultra-low CD19 expression on myeloma cells that triggers elimination by CD19 CAR-T. *Nat Commun.* 2019;10(1):3137.

72. Querques I, Mades A, Zuliani C, et al. A highly soluble Sleeping Beauty transposase improves control of gene insertion. *Nat Biotechnol.* 2019;37(12):1502-1512.

73. Monjezi R, Miskey C, Gogishvili T, et al. Enhanced CAR T-cell engineering using nonviral Sleeping Beauty transposition from minicircle vectors. *Leukemia*. 2017;31(1):186-194.

Tables and Figures:

Table 1: Top 10 burning questions for immunotherapy in MM

Questions	Assessment and perspective in 2020
Will high-risk disease become standard-risk	Potentially yes: BCMA-CAR T-cells are
in the era of novel CAR T-cell therapy?	effective in patients with high-risk
	cytogenetics, incl. double hit TP53 mutation.
Is MRD negativity the best endpoint for	Probably yes. Yet, there is an issue with
immunotherapy trials?	obtaining high quality MRD samples shortly
	after CAR T due to hypocellular BM.
	Furthermore, MRD assessment should be
	combined with functional imaging to exclude
	residual focal lesions or extramedullary disease.
Should we deliver novel immunotherapies	Probably yes. T cells from patients in early
preferentially in early treatment lines (and	disease stages and early in the treatment
what is the optimal sequence of novel	course exhibit better fitness compared to
immunotherapies)?	late stages. Determining the optimal
	sequence of immunotherapy modalities is
	an open issue, e.g.: i. What is the optimal
	interval between Dara and Elo (because
	Dara diminishes NK cells that are required
	for Elo's mode of action? ii. If one BCMA-
	targeting agent fails, can another still work?
	iii. How soon after CAR-T therapy can
	bsAbs be considered (if lymphopenia after
	lymphodepletion has not resolved)?
Is relapse from immunotherapy more	Probably no. Yet, the selective pressure of
difficult to treat?	immunotherapies on tumor and
Milling ours musicans with CAD T calls and	microenvironment has yet to be defined.
Will we cure myeloma with CAR T-cells and	Probably yes, a subgroup of patients with
T-cell engaging bispecific antibodies?	low tumor burden and favorable biology will
Will we deliver chemotherapy-free	exceptionally benefit. Probably yes. Strategies include combo of
immunotherapy combos?	anti-CD38 mAbs with ADCs or bsAbs (anti-
	BCMA or anti-GPRC5D) or combo of T cell
	redirecting therapies with IMIDs to
	reprogram and improve fitness of
	endogenous T cells.
Do we need additional diagnostics in the	Probably yes. There is a need and
era of immunotherapy?	opportunity to implement advanced
	diagnostics into MM immunotherapy to
	guide patient and antigen selection, and to
	monitor disease response as well as
	advanced genomic analyses to monitor
	clonal composition and evolution, potential

	gene or even chromosomal loss.
Will targeting more than 1 antigen on MM	Probably yes. Multi-antigen targeting is an
cells improve response rates and durability	appealing strategy to counteract antigen
of response?	down-regulation and -loss that limits the
	efficacy of BCMA-CAR T-cells in a subset of
	of patients in current clinical trials.
Is there a place for immunotherapy in early	Probably yes, but these therapies need to
asymptomatic stages of MM?	be well tolerated.
Will the COVID 19 pandemic stop the	Definitely no, but the timelines for
success of novel immunotherapies?	completion of trials, approval of novel trials
	and reimbursement may decelerate.

Figure legend

Figure 1: A) Comparison of immunotherapy treatment modalities (CAR - chimeric antigen receptor; bsAb – bispecific (T-cell engaging) antibody; ADC – antibody drug-conjugate; mAb – monoclonal antibody). **B)** Potential therapeutic sequence for newly diagnosed multiple myeloma: Debulking with anti-CD38 antibody based regimen, consolidation and induction of MRD-negativity with T-cell redirecting therapy such as CAR T-cell therapy, followed by maintenance. Grey cells – multiple myeloma cells. Light cells – T cells.

Figure 1

